

## Article

## Karyotype Analysis on Panettone Yeasts Chromosome

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## &lt;ABSTRACT&gt;

Panettone is one of the traditional sweet breads eaten in the northern part of Italy in the Christmas season. It is one of the sour breads, and is generally made by the natural starter method. It is widely known that fermentation of the dough is done by yeast and the symbiosis of the lactic acid bacterium. Many researchers have previously tried to isolate and identify the yeast and the lactic acid bacterium from the mother dough. Regarding the lactic acid bacterium, *Lactobacillus sanfranciscensis* has been isolated mainly, so far. Regarding the yeasts, *Saccharomyces exiguus*, *Saccharomyces cerevisiae*, *Candida holmii*, and *Candida humilis* have been mainly isolated. These four kinds of yeast have a difficult classification of the genus class by phenotype, and, by the identification methods by various phenotypes and genotypes by the spacer sequence of ribosomal DNA. The present conditions are that a different scientific name is referred to by each researcher. Therefore, during classification of the panettone yeast, karyotypic analysis of the yeast into these four species using the pulsed-field gel electrophoresis (PFGE) was performed as an influential taxonomic source of information. As a result, for the classification method of the genus class by the past taxonomic studies, a review was necessary for the whole, and the observed karyotype pattern suggested that they are generally divided into three groups.

Keywords: panettone, yeast, chromosome, karyotype, taxonomy

## INTRODUCTION

Panettone is one of the sweet breads made from sourdough, baked in each family in the Christmas season for more than 150 years in northern Italy<sup>1)</sup>. It features a unique and favorable sweet-sour flavor. The predominant dough-fermentable microbes, that is, yeasts and lactic acid bacteria, are considered to have originated from natural vegetable matter such as grape must, raisins, apples, figs, lemon, or orange peels, bran, and hay or horse dung which are added to the wheat flour dough to prepare the starter “mother dough”<sup>2)</sup>. Few studies have

been performed of the characterization of lactobacilli and yeasts isolated from panettone sour doughs<sup>3-6)</sup>.

*Torulopsis holmii* was the first to be isolated from panettone sourdough<sup>7)</sup>. Later, those yeasts were ascribed to *Saccharomyces exiguus*, *Saccharomyces cerevisiae* and *Candida stellate*<sup>8)</sup>. The followed study<sup>9)</sup> showed yeast strains belonging to *Candida holmii*, which were the asexual form of *Saccharomyces exiguus* and *Saccharomyces cerevisiae*. A recent taxonomic study<sup>2)</sup> according to the sequences of the internal transcribed spacers between 18S and 26S rDNA indicated that the identification system based on

phenotypic analysis proved to be unreliable to identify yeasts from sourdough, and showed that *Candida humilis* was the predominant species, whereas the remaining strains were related to the *Saccharomyces cerevisiae* stricto group. The aim of this study is to provide influential and effective information on the past taxonomic studies of panettone yeasts through the use of chromosome karyotype, and to reconsider the classification.

## MATERIALS AND METHODS

### **Yeast strains**

*Saccharomyces cerevisiae* S288C (type culture) was obtained from ATCC (American Type Culture Collection, 10801 University Boulevard Manassas, VA20110, USA), and used as the control strain for the pulsed-field gel electrophoresis. Following strains were the same strains that were found in Panettone mother doughs, and they were obtained from NBRC (Nite Biological Resource Center, 2-49-10 Nishihara, Shibuyaku, Tokyo, Japan). These strains have been identified by means of phenotypic methods.

*Candida holmii* NBRC 0660

*Candida holmii* NBRC 1629

*Candida humilis* NBRC 10280

*Saccharomyces exiguus* NBRC0215

*Saccharomyces exiguus* NBRC0271

*Saccharomyces exiguus* NBRC0956

*Saccharomyces exiguus* NBRC1128

*Saccharomyces exiguus* NBRC1141

*Saccharomyces exiguus* NBRC1142

*Saccharomyces exiguus* NBRC1169

*Saccharomyces exiguus* NBRC1170

*Saccharomyces exiguus* NBRC1617

*Saccharomyces exiguus* NBRC10181

The following strains were isolated from a panettone sourdough and were obtained from Professor Roberto Foschino, Department of Food Science and Microbiology (DiSTAM), Faculty of Agricultural, University of Milan (Via Celoria, 2-20133 Milano, Italy). These strains were identified by genotypic method according to the rDNA spacer sequence<sup>2)</sup>.

*Candida humilis* RM8

*Candida humilis* #4

*Candida humilis* LGAT

*Candida humilis* #8

### **Preparation of agarose embedded yeast DNA**

A single colony on a YPD (1% yeast extract, 2% peptone, 2% glucose, pH5.8) agar plate was inoculated into 2 ml a YPD broth. After being grown for 20hrs to a stationary growth phase, the cells were collected by centrifugation at  $5,000 \times g$ , 5min, at 25°C. The supernatant was decanted and the cells were re-suspended in 1ml SE (75mM NaCl, 25mM pH7.4EDTA). Cells were collected again by centrifugation at  $5,000 \times g$ , 5min, at 25°C. 50  $\mu$ l of SE and 150  $\mu$ l of agarose suspension buffer (ASB) which was equilibrated to 55°C in a water bath were added. ASB was freshly prepared in each experiment. 200 $\mu$ l of ASB was mixed with the following four solutions, 500 $\mu$ l of 3.5% low melt agarose (BRL, Gaithersburg, MD, USA) which was equilibrated to 55°C in a water bath after being autoclaved in SE for 1min at 121°C, 20 $\mu$ l of 1M DTT (filter-sterilized after dissolved in water), 10 $\mu$ l of 1mg/ml Zymolyase (Kirin 100T, Kirin Brewery Co. Ltd, Tokyo, Japan) suspended in sterile water, and 470  $\mu$ l of SE. After immediately combining the cell mixture, it was transferred to the mixture plug mold and solidified for 8min at -20°C. Each solidified agarose plug was pushed into a well of 20 well micro plate and 80  $\mu$ l of 1M DTT (filter-sterilized after dissolved in water), 40  $\mu$ l of 1mg/ml Zymolyase (Kirin 100T, Kirin Brewery Co. Ltd, Tokyo, Japan) suspended in sterile water, 1880  $\mu$ l of SE were added into each well. After incubated for 1hr at 37°C, each agarose plug was washed with ES (0.5M pH9.5EDTA, 1% sodium sarcosinate) twice for 10min. 1ml ES and 50  $\mu$ l of proteinase K (Merk, Darmstadt, FRG) solution (10mg/ml in 50% glycerol) was added to each well and incubated at 55°C for 20 hrs. After proteinase K treatment, each agarose plugs were washed with  $1 \times$  TE (20mM Tris-HCl, 1mM EDTA, pH7.4) twice for 20min and stored at 4°C until

use.

### ***Pulse-field gel electrophoresis (PFGE)***

CHEF (Contour-clamped homogenous electric field) gel electrophoresis was performed as described elsewhere<sup>10)</sup>, using a CHEF-DR II system (Nippon Bio-Rad Laboratories, Tokyo, Japan). The 1% agarose gels in 0.5×TBE (45mM Tris-borate, 45mM boric acid, 1mM pH8.0 EDTA) were prepared by pouring 100ml agarose (Agarose NA, Pharmacia AB, Uppsala, Sweden) into a 12×12cm frame. Electrophoresis was carried out in 0.5×TBE as running buffer at a constant temperature of 14°C, which was maintained by re-circulation of the buffer through a heat exchanger, at a constant voltage of 170V for 20hrs. The stepped switching interval was 60sec for the first 12hr and 90sec for the next 8hrs. The gel was stained with 0.0025% SYBR Green solution (TAKARA BIO Inc., Tokyo, Japan) for 1hr with moderate shaking in darkness. The resulting gel images were captured by Chemi-Imager4400 (Alpha Innotech corporation, San Leandro, CA, USA).

### ***Cluster analysis***

Chromosome karyotype was treated with Fingerprinting II Software ver.3 (Bio-Rad Laboratories, 2000 Alfred Nobel Drive, Hercules, CA94547, USA) which is designed to analyze banding patterns and fingerprints from I-D gels, chromatograms, and density curves. Manual grouping was performed on the result by the chromosome number, then the phylogenetic tree was estimated to obtain the relationships of yeast strains.

## **RESULTS AND DISCUSSION**

### ***Analysis of pulsed-field gel electrophoresis***

The CHEF electrophoretic pattern of all the strains was shown in Figure 1. Different karyotypes were observed among tested strains of *S. exiguus*, indicating that phenotypic classification may not be correct. The same

result was shown in two other strains, *C. holmii* and *C. humilis*. Since the wild yeast is usually polyploid, the exact chromosome number is not clear. But the chromosome number can be estimated roughly by means of a combination of the band numbers and the intensity of the band. When looking at the smaller sized chromosome, the second and the third chromosomes of *S. exiguus* NBRC1142 exist closer for *S. exiguus* NBRC0956. And those two chromosomes seem to overlap for *S. exiguus* NBRC1169. Therefore these three strains are very close evolutionally. *S. exiguus* NBRC0271 showed similar karyotype to *S. exiguus* NBRC1128, *S. exiguus* NBRC1141, and *S. exiguus* NBRC1170 (Group A). Group A is very similar to *S. cerevisiae*. *S. exiguus* NBRC0956 showed a similar karyotype to *S. exiguus* NBRC1142 and *S. exiguus* NBRC1169 (Group B).

*S. exiguus* NBRC0215, *S. exiguus* NBRC1617 and *S. exiguus* NBRC10181 were different to each other from Group A and B. Therefore five different patterns of karyotypes were found among *S. exiguus* strains.

The karyotype of *C. holmii* NBRC0660 is different from *C. holmii* NBRC1629. The karyotype of *C. holmii* NBRC0660 is very similar to *S. exiguus* NBRC0956 (Group B) though the chromosome number is one or two excess in *C. holmii* NBRC0660 compared with *S. exiguus* NBRC0956 (Group B). On the other hand, *C. holmii* NBRC1629 showed a similar karyotype to *S. exiguus* NBRC1170 (Group A).

There were two groups for *C. humilis*. *C. humilis* NBRC10280 showed a very similar karyotype to *C. humilis* #4 and *C. humilis* RM8 (Group "a"). Group "a" showed a similar karyotype to *S. exiguus* NBRC0956 (Group B). *C. humilis* #8 showed a very similar karyotype to *C. humilis* LGAT (Group "b"). Since *C. holmii* is the asexual form of *S. exiguus* and *S. cerevisiae*<sup>9)</sup>, the karyotype should resemble each other. Actually, the karyotype of Group "a" is similar to the karyotype of Group B. And the karyotype of Group "b" is similar to the karyotype of Group A. As the result mentioned above, it is

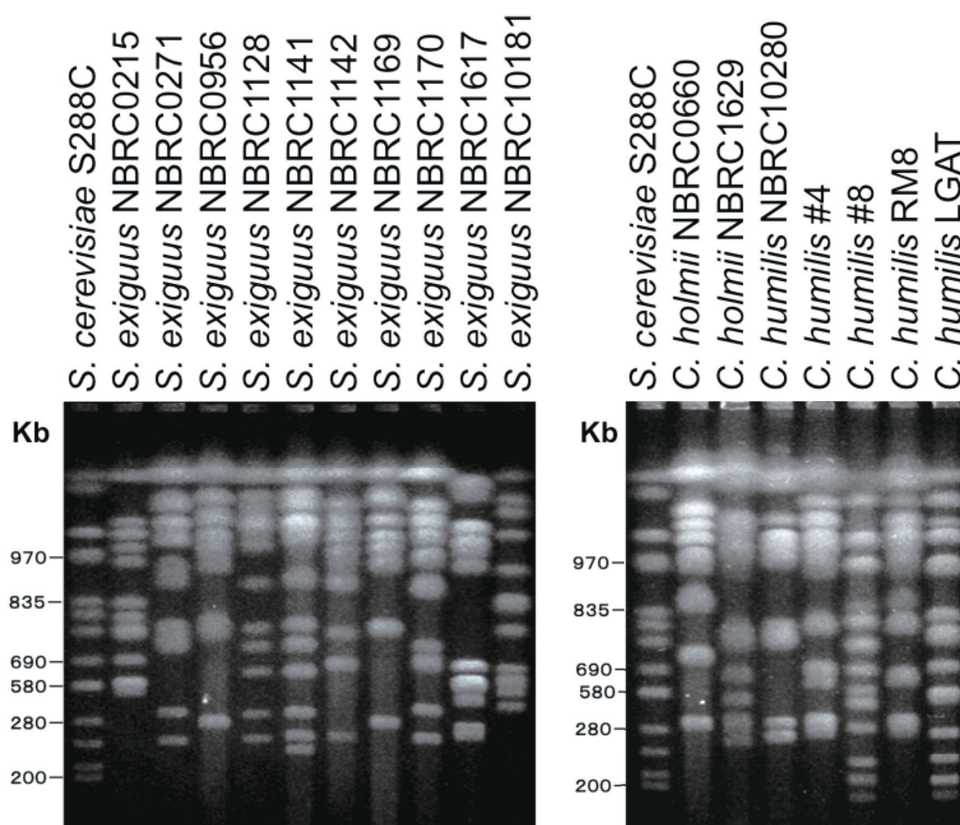


Figure 1. The result of PFGE analysis on yeast strains found in panettone mother dough. The chromosome of *Saccharomyces cerevisiae* S288C was used as a size-marker.

suggested that the karyotype was classified roughly into two groups and one other group, totally three groups.

### ***Relationships of yeast strains***

Generally, the phylogenetic tree of yeast is drawn by means of rDNA sequences<sup>11, 12</sup>. When creatures are classified, the chromosome number is often used as an effective means. In the case of yeast, the number of chromosomes can be ascertained by pulsed-field gel electrophoresis (PFGE), but there are few examples which made a phylogenetic tree among yeast strains by their karyotypes. Electrophoretic patterns of microbes DNA cleaved with a restriction enzyme can be analyzed by Fingerprinting II Software ver.3 (Bio-Rad Laboratories)<sup>13</sup>. The software is designed to analyze banding patterns and fingerprints from I-D gels, chromatograms, and density curves. This software is the only a tool at present in the

world to analyze the electrophoretic pattern of PFGE. Because this software can't consider the chromosome number for cluster analysis, a phylogenetic tree for yeast strains is hard to draw. Thus, a manual grouping was performed on the result of the software by the chromosome number, as the result of which the phylogenetic tree was estimated. The phylogenetic tree drawn according to the electrophoretic patterns of Figure 1 is shown in Figure 2.

As previously mentioned, Figure 2 demonstrates that strains were classified into two big groups and one other. The strains of *S. exiguus* were divided into five categories, and, as for this classification, it is strongly suggested that reconsideration is necessary. *C. humilis* was divided into two groups, and one group was considerably similar to *S. cerevisiae*, and the karyotype resembles it, with only one or two chromosome numbers being different.

Foschino *et al.*<sup>2</sup> performed a comparison of



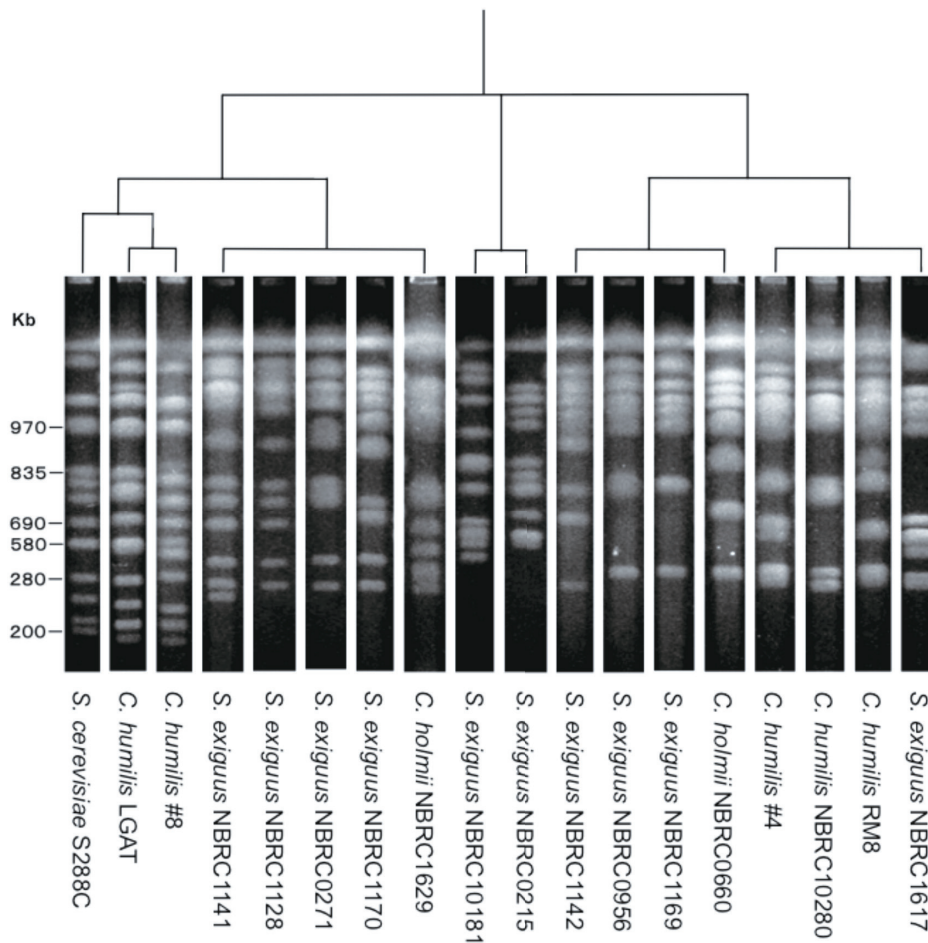


Figure 2. Relationships of selected yeast strains found in panettone mother dough.

classifying the yeast strains found in Panettone mother dough between the recent genotypic method<sup>11)</sup> (rDNA analysis) and the traditional phenotypic methods<sup>14, 15)</sup>. They showed that a big discrepancy occurred in the classification by the phenotype according to the difference in the analytical technique. Also, big difference was observed in the classification between the recent taxonomic method whose analysis is based on rDNA sequence and the past taxonomic method whose analysis is based on phenotype, that is, physiological characteristics and assimilation capabilities based on different carbon sources. In their study, among the three phenotypic methods, surprisingly there was not even one case that matched a result accorded. As a matter of course, the phenotypic methods did not match with the recent genotypic method

either. According to the recent rDNA analysis, they reclassified Panettone yeasts into three species, that is, *S. cerevisiae*, *C. humilis*, *S. pastorianus*.

By our result using the karyotype analysis, the Panettone yeasts were classified into at least five categories, and if *S. cerevisiae* is included into the Panettone yeasts as shown by Foschino *et al.*<sup>2)</sup>, the yeasts were classified into six categories. The recent taxonomic method for yeast is not taking into account the chromosome karyotype for yeast classification. But our result lead us to strongly urge that the classification method of the genus class of yeast should be reviewed on the standing point of considering the balance among the chromosome karyotype, rDNA sequence, and phenotypic feature.

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## パネットーネ酵母染色体の核型分析

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### <要 旨>

パネットーネとはイタリア北部でクリスマスシーズンに食される伝統的なスイートブレッドのひとつである。パンの分類上はサワーブレッドのひとつであり、自然種製法によって作られる。酵母と乳酸菌の共生によってパン生地が発酵がなされることが広く知られている。これまで多くの研究者が母種から酵母と乳酸菌の分離・同定を試みてきている。乳酸菌については、主に、*Lactobacillus sanfranciscencis*が分離されてきている。酵母については、主なものとして*Saccharomyces exiguus*、*Saccharomyces cerevisiae*、*Candida holmii*、*Candida humilis*が分離されてきている。これら4種の酵母は表現型による属種の分類が難しく、各種表現型やリボソームDNAのスペーサー配列による遺伝子型による同定方法などによって、研究者によってそれぞれ異なる学名が付されているのが現状である。そこで、パネットーネ酵母の分類上、有力な情報源としてこれら4種の酵母の核型解析をパルスフィールドゲル電気泳動（PFGE）によって行った。その結果、核型から観ると、これまでの研究による属種の分類方法は全体に見直しが必要であり、大きく3群に分けられることが示唆された。

キーワード：パネットーネ、酵母、染色体、核型分析、分類